

Towards schizozygine: synthesis of 15 α -hydroxystrempeliopine

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Abstract—15 α -Hydroxystrempeliopine (**6**), a model hexacyclic base for our planned total synthesis of schizozygine (**2**), has been synthesized using a Zn-mediated reductive rearrangement of indolenine **14** as the key step. The structure and relative stereochemistry of **6** were assigned by a combination of COSY, NOESY, HSQC and HMBC correlations.
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Schizozyganes constitute a small group of post-secodine hexacyclic alkaloids, which have been known for more than 40 years.^{1–3} With the exception of (–)-strempeliopine (**1**) (Fig. 1), an alkaloid of the Cuban species *Strempeliopsis strempelioides* K. Schum.,⁴ all have been isolated from the East-African monotypic shrub *Schizozygia coffaeoides* (Boj.) Baill.⁵ This plant has been used as a traditional medicine for skin diseases;⁶ leaf extracts show high antifungal and antimicrobial activity.⁷ (+)-Schizozygine (**2**) is the major alkaloid accompanied among others, by the minor bases 3–5 (Fig. 2).⁸

Schizozygane alkaloids are attractive synthetic targets due to their interesting skeleton and pharmaceutical properties. While the synthesis of *seco*-schizozygane/vallesamidine bases has been described repeatedly,^{9–14} only one synthesis of the parent base of the group, (–)-strempeliopine (**1**), has been reported¹⁵ to date. This is based on a reductive rearrangement of an indolenine.^{9,10}

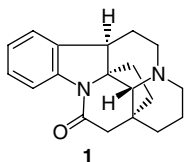


Figure 1. (–)-Strempeliopine.

Keywords: Indole alkaloids; Schizozygane alkaloids; Strempeliopine; Alkaloid synthesis; Indolenine; Reductive rearrangement.

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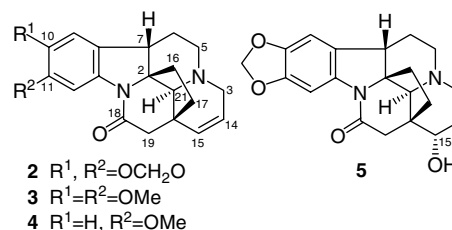
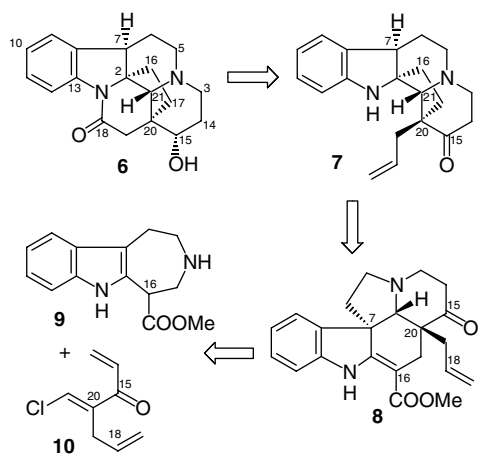


Figure 2. Selected alkaloids from *Schizozygia coffaeoides* (Boj.) Baill.

Recently, a synthesis of the biogenetically related alkaloid isoschizogamine was also described.¹⁶

Renner has proposed² that the 14,15-unsaturation in bases 2–4 might be derived from the corresponding 15-ketone. However, the implementation of such a strategy in the synthesis of schizozygine (**2**) would require proof of compatibility of the carbonyl group with the anticipated reaction conditions. We now report on the successful realization of such an approach and describe a synthesis of 15 α -hydroxystrempeliopine (**6**).¹⁷

Thus, it was expected that the base **6** might be synthesized by reductive rearrangement of an indolenine with suitable functionalization at C-18. An allyl group at C-20 has already proved to be an effective precursor for the lactam by ozonolytic oxidative cleavage.^{15,18} The introduction of oxygen at C-15 would result in intermediate **7**, which could be derived from β -anilinoacrylate **8** (Scheme 1). This should be accessible by application of Kuehne's efficient protocol¹⁹ starting from indoloazepine **9**.²⁰ 1-Chloro-2-allyl-1,4-pentadienone (**10**) was designed as the second building block for the preparation of **8**.

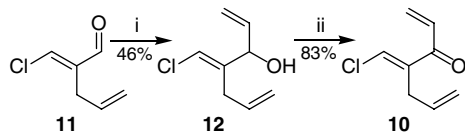
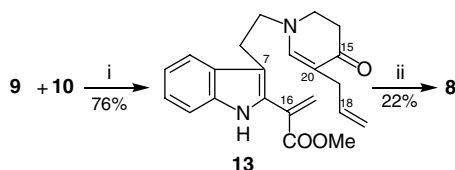
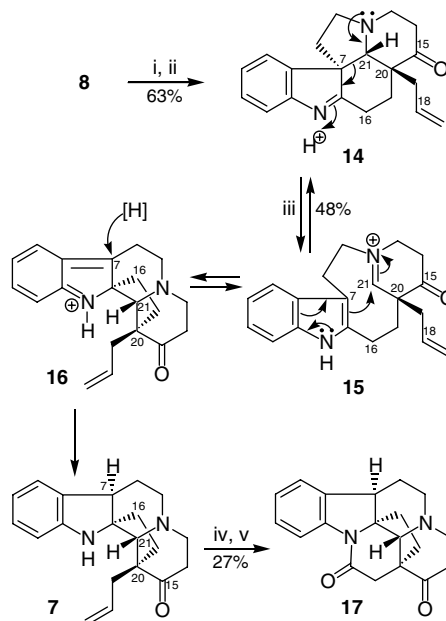


Scheme 1. Retrosynthetic analysis.

Dienone **10** was prepared from 2-allyl-3-chloroacrolein²¹ **11** by Grignard reaction with vinylmagnesium bromide at 0 °C in THF and subsequent oxidation of the resulting sensitive alcohol **12** by Dess–Martin periodinane (Scheme 2).

Condensation of the dienone **10** with indoloazepine **9** in methanol at room temperature for 20 min afforded an unstable (rapid dimerization) secodine derivative **13** (76% yield), which was immediately subjected to a [4+2] cycloaddition reaction in boiling toluene (Scheme 3). The desired 18-methylene-15-oxovincadifformine (**8**) was thus obtained in a 22% yield.

Alkaline hydrolysis of ester **8** followed by decarboxylation of the acid in boiling benzene gave the indolenine **14** in 63% yield (Scheme 4). Zinc-mediated reductive rearrangement of **14** in the presence of catalytic CuSO₄·5H₂O in hot acetic acid^{10,15} proceeded presumably via the intermediary iminium ions **15** and **16**,²² and led to the formation, in 48% yield, of the 2,2,3-trisubstituted indoline **7**, which was formylated by the mixed-anhydride method.²³

Scheme 2. Synthesis of dienone **10**. Reagents and conditions: (i) vinylmagnesium bromide, THF, 0 °C; (ii) Dess–Martin periodinane, dichloromethane, 4 °C to rt.Scheme 3. Synthesis of 18-methylene-15-oxovincadifformine (**8**). Reagents and conditions: (i) MeOH, rt, 20 min, rapid isolation of **13** under cooling; (ii) toluene, reflux, 20 h.Scheme 4. Synthesis of 15-oxostrempeliopine (**17**). Reagents and conditions: (i) KOH, EtOH, reflux, 4 h; (ii) benzene, reflux, 3 h; (iii) Zn, CuSO₄·5H₂O (cat.), AcOH, 105 °C, 5 h; (iv) HCOOH, Ac₂O, rt, overnight; (v) O₃, MeOH, 1 M H₂SO₄, 0 °C then 30% H₂O₂, 18 h.

Synthesis of 15-oxostrempeliopine (**17**) was completed (27%) by gentle ozonolysis in a mixture of 1 M sulfuric acid/methanol at 0 °C followed by treatment with hydrogen peroxide. Finally, smooth reduction of ketone **17** proved to be highly diastereoselective even with

Table 1. ¹H and ¹³C NMR data of **6** in CDCl₃ at 300 K^a

	δ _H	δ _C	HMBC (¹ H)
2		71.9	6β, 7α, 16, 17α
3α	2.90 (1H, ddd, 11.6, 4.0, 2.7)	52.7	5β, 14, 21β
3β	2.17 (1H, btd, 11.7, 3.3)		
5α	2.97 (1H, ddd, 11.3, 7.9, 5.9)	49.7	3α, 6, 7α, 21β
5β	2.23 (1H, dt, 11.4, 6.0)		
6α	2.11 (1H, bdq, 14.2, 6.2)	26.2	7α, 5β, 5α
6β	1.97 (1H, dtd, 14.2, 7.9, 5.9)		
7α	3.25 (1H, bt, 7.1)	41.6	5, 6α, 21α
8		132.8	12, 10, 6, 7α
9	7.17 (1H, d, 7.4)	123.6	7α
10	7.07 (1H, t, 7.4)	124.0	12
11	7.23 (1H, t, 7.7)	128.1	9, 7α
12	8.06 (1H, d, 8.0)	115.9	10
13		142.0	12, 11, 7α
14α	1.89 (1H, m)	31.0	3
14β	1.83 (1H, m)		
15β	3.67 (1H, dd, 10.6, 5.4)	74.5	3β, 19
16a	2.33 (1H, m)	37.8	7α, 17β, 21β
16b	2.31 (1H, m)		
17a	2.29 (1H, m)	27.8	15β, 16, 19, 21β
17b	1.63 (1H, m)		
18		168.9	19
19α	2.95 (1H, d, 18.2)	48.2	15β
19β	2.56 (1H, dd, 18.2, 2.4)		
20		48.5	17, 19
21β	2.09 (1H, s)	68.9	3α, 5β, 7α, 16, 19

^a Data from Bruker Avance, measured at 500.13 MHz and 125.77 MHz, respectively; for atom numbering, see structure **6**, Scheme 1.

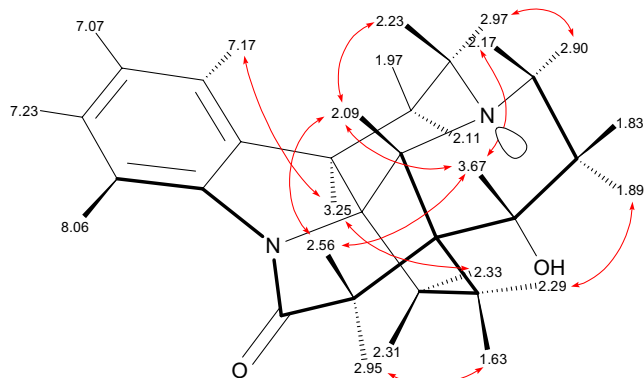


Figure 3. Observed non-trivial NOE interactions in base 6.

sodium borohydride in ethanol and provided a 90% yield of the desired base 6.

The base 6 showed a pseudomolecular ion peak at m/z 310 (M^+), which corresponded to the molecular formula $C_{19}H_{22}N_2O_2$. The structure and stereochemistry was assigned on the basis of both 1D and 2D 1H and ^{13}C NMR experiments (Table 1 and Fig. 3).

The 1H NMR spectrum displayed characteristic proton signals at δ 3.25 ppm (H-7) and 2.09 ppm for H-21, and the 1H – 1H COSY spectrum showed a diagnostic long-range coupling (W) between H-19 at δ 2.56 ppm and H-17a at δ 2.29 ppm ($J = 2.4$ Hz). The relative configuration at C-15 stems from the fact the signal at δ 3.67 ppm showed as a cross peak in the NOESY spectrum with H-3 β at δ 2.17 ppm as well as with H-21. Moreover, H-15 appears as a doublet of doublets ($J = 10.6$ and 5.4 Hz), while the H-15 signal in the 1H NMR spectrum of epimeric natural (+)- α -schizozogol (5) appears as a triplet with $J \sim 3.0$ Hz.²⁴ The base is thus 15 α -hydroxystrempeliopine (6) with an equatorial hydroxy group. Not surprisingly, alcohol 6 failed to afford any olefine on attempted dehydration through tosylation.

The application of this strategy to a total synthesis of schizozogine (1) is underway. The results will be published in due course.

References and notes

1. Renner, U.; Kernweisz, P. *Experientia* **1963**, *19*, 244–246.
2. Renner, U. *Lloydia* **1964**, *27*, 406–415.

3. Renner, U.; Fritz, H. *Helv. Chim. Acta* **1965**, *27*, 308–317.
4. Laguna, A.; Novotný, L.; Dolejš, L.; Buděšínský, M. *Planta Med.* **1984**, *50*, 285–288.
5. Taylor, W. I. In *The Alkaloids, Chemistry and Physiology*; Manske, R. H. F., Ed.; Academic Press: New York, 1974; Vol. 11, pp 137–142.
6. Omino, E. A.; Kokwaro, J. O. *J. Ethnopharmacol.* **1993**, *40*, 167–180.
7. (a) Kariba, R. M.; Siboe, G. M.; Dossaji, S. F. *J. Ethnopharmacol.* **2001**, *74*, 41–44; (b) Kariba, R. M.; Houghton, P. J.; Yenesew, A. *J. Nat. Prod.* **2002**, *65*, 566–569.
8. LeMen, J.; Taylor, W. I. *Experientia* **1965**, *21*, 508–511.
9. Maupérin, P.; Lévy, J.; LeMen, J. *Tetrahedron Lett.* **1971**, 999–1002.
10. Lévy, J.; Maupérin, P.; de Maidreville, M. D.; LeMen, J. *Tetrahedron Lett.* **1971**, 1003–1006.
11. (a) Dickman, D. A.; Heathcock, C. H. *J. Am. Chem. Soc.* **1989**, *111*, 1528–1530; (b) Heathcock, C. H.; Norman, M. H. Dickman, D. A. *J. Org. Chem.* **1990**, *55*, 798–811.
12. Costa, P. R. R.; Castro, R. N.; Farias, F. M. C.; Antunes, O. A. C.; Bergter, L. *Tetrahedron: Asymmetry* **1993**, *4*, 1499–1500.
13. Padwa, A.; Haring, S. R.; Semones, M. A. *J. Org. Chem.* **1998**, *63*, 44–54.
14. (a) Tanino, H.; Fukuishi, K.; Ushiyama, M.; Okada, K. *Tetrahedron Lett.* **2002**, *43*, 2385–2388; (b) Tanino, H.; Fukuishi, K.; Ushiyama, M.; Okada, K. *Tetrahedron* **2004**, *60*, 3273–3282.
15. (a) Hájiček, J.; Trojáněk, J. *Tetrahedron Lett.* **1981**, *22*, 2927–2928; (b) Hájiček, J.; Trojáněk, J. *Tetrahedron Lett.* **1982**, *23*, 365–368; (c) Hájiček, J.; Trojáněk, J. *Collect. Czech. Chem. Commun.* **1986**, *51*, 1731–1742.
16. Hubbs, J. L.; Heathcock, C. H. *Org. Lett.* **1999**, *1*, 1315–1317.
17. Modified ^{15c} biogenetic numbering is used in this paper: LeMen, J.; Taylor, W. I. *Experientia* **1965**, *21*, 508–509.
18. Hájiček, J.; Trojáněk, J. *Collect. Czech. Chem. Commun.* **1982**, *47*, 2448–2453.
19. Kuehne, M. E.; Bornmann, W. G.; Earley, W. G.; Marko, I. *J. Org. Chem.* **1986**, *51*, 2913–2927.
20. Kuehne, M. E.; Bohnert, J. C.; Bornmann, W. G.; Kirkemo, C. L.; Kuehne, S. E.; Seaton, P. J.; Sebovitz, T. C. *J. Org. Chem.* **1985**, *50*, 919–923.
21. (a) Arnold, Z.; Šorm, F. *Chem. Listy* **1957**, *51*, 1082–1087; (b) Arnold, Z.; Žemlička, J. *Collect. Czech. Chem. Commun.* **1959**, *24*, 2378–2385.
22. Palmisano, G.; Danieli, B.; Lesma, G.; Trupiano, F.; Pilati, T. *J. Org. Chem.* **1988**, *53*, 1056–1064.
23. Formylation both changes polarity and thus facilitates separation of related minor impurities, and protects indoline nitrogen in the subsequent step.^{15c}
24. Hájiček, J. Unpublished results.